A Review of Genetic Alteration Correlated with Primary and Acquired Resistance to EGFR–TKIs in EGFR–Mutant Non–Small Cell Lung Cancer

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Abstract:

Epidermal growth factor receptor tyrosine kinase inhibitors (EGFR-TKIs) significantly enhance both survival and quality of life for non-small cell lung cancer (NSCLC) patients with EGFR mutations. Nevertheless, an increasing number of patients exhibit primary or acquired resistance. Currently, discoveries of concurrent genetic alterations in EGFR and other genes besides EGFR have been made. Several studies have demonstrated an association between these alterations and resistance to EGFR-TKI. Therefore, it is crucial to gain more understanding of gene alterations and their impact on the effectiveness of EGFR-TKI treatment. In this review, concurrent gene alterations that can predict primary and acquired resistance to EGFR-TKIs in advanced NSCLC are discussed. This may assist clinicians in selecting the appropriate treatment for patients who have failed to respond to EGFR-TKIs.

Keywords: acquired resistance, EGFR-TKIs, genetic alteration, non-small cell lung cancer, primary resistance

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Introduction

Lung cancer is the second most frequently diagnosed cancer globally, with an estimated age-standardized rate (ASR) of 22.4. It is also the primary cause of cancerrelated deaths, with an ASR of 18.0¹. In Thailand, lung cancer is the third most common type of cancer, having an ASR of 18.9, and it is the second leading cause of cancer-related deaths, with an ASR of 15.9². Non-smallcell lung carcinoma (NSCLC) is the predominant histologic type, accounting for 85% of lung cancer cases³. Over time, driver mutations in NSCLC have been discovered, enabling the development of personalized targeted treatments that enhance response rates, minimize toxicity, and improve the quality of life for patients⁴. Epidermal growth factor receptors (EGFR) are prevalent driver genes in NSCLC, and are found in 40-60% of Asian populations and 10% of Western populations⁴.Overexpression or mutation of the EGFR gene results in aberrant cell growth, differentiation, survival, enhanced formation of blood vessels, and the migration of cancer cells. EGFR tyrosine kinase inhibitors (EGFR-TKIs) can substantially extend the median survival of patients with advanced lung cancer who have EGFRsensitive mutations⁵. Additionally, it is associated with a high response rate, ranging from 60% to 80%, and results in fewer side effects compared to platinum-based chemotherapy^{6,7}.

Recently, different EGFR-TKI types, including firstgeneration EGFR-TKIs (gefitinib, erlotinib, and icotinib) and second-generation EGFR-TKIs (afatinib and dacomitinib) have been authorized for treating patients with EGFRmutant NSCLC⁸; however, a large number of patients have disease relapse due to treatment resistance^{9,10}. Some patients treated with gefitinib, erlotinib, or afatinib are susceptible to primary resistance, but most eventually will acquire complete resistance¹¹. Several studies have shown that more than 20% of patients contend with early relapse within 6 months¹²⁻¹⁵. Herein, we reviewed the clinical impact of treatment, focusing on overall response rate (ORR), overall survival (OS), and progression-free survival (PFS). We also reviewed the studies that evaluated concurrent genomic alterations related to resistance in EGFR-mutated NSCLC after EGFR-TKI treatment.

EGFR mutation

The human EGFR gene is located on the short arm of chromosome 7 (7p11.2). It spans about 200 kb, which contains 28 exons and encodes a protein of 1,210 amino acids¹⁶. EGFR mutations are classified, according to the American College of Medical Genetics and Genomics (ACMG), into 3 categories: pathogenic, likely pathogenic, and benign, as shown in Figure 1¹⁷. Exon 19 has the highest mutation frequency at 48.8%, with the majority being pathogenic mutations. Of all the mutations, 18 were deletions, which may be either pathogenic or benign. Exon 21 has the second highest number of pathogenic mutations, totaling 30.95%. Exon 20 has a mutation rate of 14.28%, including 6 pathogenic mutations and 3 benign mutations. Exon 18 has a mutation rate of 5.9%, with 3 mutations classified as pathogenic, and one as likely pathogenic¹⁸. Drug-sensitive mutations include point mutations in exon 18 (G719X), point mutations in exon 21 (L858R), and in-frame deletions in exon 19 (Del19). Drug-resistant mutations are found in exon 20 (T790M)¹⁹.

Frequency of EGFR mutations in patients with NSCLC

EGFR is the most prevalent driver mutation in NSCLC, frequently found in ADC (40–60%)²⁰. EGFR mutations are rare in SCC, having a prevalence of less than 3%²¹. The lack of reported mutations may restrict the beneficial effects of EGFR–TKIs in treating lung cancer patients with SCC. By geographic region, the prevalence of EGFR mutation is highest in Asia–Pacific, ranging from 20% to 76%, with an average frequency of 47%. Vietnam

has the greatest EGFR mutation frequency in the Asia-Pacific region at 64%, while the Republic of Korea stands at 43%. The frequency of EGFR mutation in Thailand is 54%. In Europe, the overall frequency of EGFR mutation is 15%, the lowest among other geographic regions. Within this region, the EGFR mutation frequency was highest in Turkey at 37%, and lowest in Austria at 7%, as shown in Table 1²⁰.

EGFR Tyrosine Kinase Inhibitors (EGFR-TKIs)

EGFR-TKIs are small molecules that prevent the binding of adenosine triphosphates (ATP) to the tyrosine

kinase domain of the EGFR within cells. This inhibition affects intracellular signaling, thereby reducing cell division and growth²². The EGFR-TKIs are categorized into 3 generations, based on the order of their development.

First-generation EGFR-TKIs

First-generation EGFR-TKIs, such as erlotinib, gefitinib and icotinib, are quinazoline-derived drugs that bind to EGFR as a reversible inhibitor. It competes with ATP for binding at the active site of the tyrosine kinase domain through non-covalent interactions²³. These drugs have strong inhibitory effects on activating EGFR mutations,

Table 1 Frequency of EGFR mutations in patients with NSCLC of ADC (Modified from Midha et al. 2015)²⁰

| Country | EGFR mutati | on frequency |
|------------------------------|-------------------------------------|-----------------------------------|
| | Overall EGFR mutation frequency (%) | EGFR mutation frequency range (%) |
| Asia-Pacific: Overall | 47 | 20-76 |
| China | 48 | 27–66 |
| Japan | 45 | 21-68 |
| Malaysia | 45 | 39–47 |
| Philippines | 52 | N/A |
| Korea | 43 | 20-56 |
| Thailand | 54 | N/A |
| Vietnam | 64 | N/A |
| Europe: Overall | <u>15</u> | <u>6-41</u> |
| Greece | 22 | 20–22 |
| Italy | 14 | 10–33 |
| The Netherlands | 15 | 11–20 |
| Poland | 13 | 11–14 |
| Russia | 22 | 20–31 |
| Slovakia | 20 | N/A |
| Turkey | 37 | 22-41 |
| United Kingdom | 12 | N/A |
| North America: Overall | <u>22</u> | <u>3-42</u> |
| Canada | 14 | 14 |
| United States of America | 23 | 3-42 |
| Indian subcontinent: Overall | <u>26</u> | <u>22–27</u> |
| India | 26 | 22–26 |
| Pakistan | 27 | N/A |
| South America: Overall | <u>36</u> | <u>9–67</u> |
| Argentina | 19 | N/A |
| Brazil | 28 | 9–34 |
| Peru | 67 | N/A |

ADC=denocarcinoma, EGFR=epidermal growth factor receptor, NSCLC=non-small cell lung cancer

either exon 19 deletions or exon 21 L858R²⁴. However, they are only effective in the short term due to the occurrence of acquired resistance caused by mutations within the drug-binding site²⁴.

Second-generation EGFR-TKIs

Second-generation EGFR-TKIs, including afatinib, dacomitinib and neratinib, are quinazoline-based drugs that act as irreversible inhibitors by binding to EGFR. The development of these drugs is intended to overcome the acquired resistance resulting from the failure of first-generation EGFR-TKIs. The mechanisms of action of second-generation EGFR-TKIs differ from those of first-generation EGFR-TKIs. The drugs exhibit an affinity via covalent bonding to cysteine residues within the tyrosine kinase domain²³.

Third-generation EGFR-TKIs

First and second-generation EGFR-TKIs have high efficacy, although resistance often develops within 10–14 months after treatment. EGFR T790M is the most common mutation associated with acquired resistance, which is present in 50–70% of patients⁶. Third-generation EGFR-TKIs are aminopyrimidine-based drugs that act as irreversible inhibitors by binding to EGFR. Osimertinib is currently recommended as a treatment option for NSCLC patients who have developed resistance to first- or second-generation EGFR-TKIs, as a second- or thirdline therapy²³.

Clinical studies regarding EGFR-TKIs treatments in advanced NSCLC patients

Herein, we have retrieved the study demonstrating objective response rate (ORR), overall survival (OS), and progression-free survival time (PFS) in various populations of advanced NSCLC patients following EGFR-TKI. As shown in Table 2, there was a total of 1,881 cases of patients harboring EGFR-activating mutations (deletion 19, G719X, L858R, and L861Q). All patients received either first-generation EGFR-TKIs (gefitinib and erlotinib) or second- generation (afatinib) EGFR-TKIs, and adenocarcinoma (ADC) was diagnosed in almost all cases (99.57%; 1,873/1,881 cases).

The ORR, which is the proportion of patients that achieved a response, was comparable for 3 EGFR-TKIs: erlotinib (53.8–78.0%), gefitinib (56.0–62.5%), and afatinib (50.0–70.0%). The median OS was 20.4 months for erlotinib, ranging from 17.0 to 25.0 months for gefitinib, and from 23.1 to 27.9 months for afatinib. The PFS, which is calculated as the interval between the initiation of treatment and progression or death, was found to be 11.2–12.1 months for erlotinib, 8.0–12.1 months for gefitinib, and 10.3–16.4 months for afatinib.

Furthermore, we have shown in this review a subset of patient recurrence of various durations. PFS was categorized into 3 time periods: less than 3 months, 3 to 6 months, and more than 6 months (Table 2). Most patients usually show relapse after treatment over 6 months: ranging from 53.6% to 80.3% of the total patients. Nevertheless, a subset of patients developed the disease within 6 months of undergoing treatment. This varied among patients from 19.7% to 46.4%. These findings suggest a pattern of patients having developed acquired, early resistance to treatment or not responding to it (referred to as primary resistance).

Resistance to EGFR-TKIs

Although EGFR-TKIs are presented as a preferred method of lung cancer treatment compared to traditional chemotherapy, some patients develop resistance after exposure to the drugs. Resistance to EGFR-TKI is clinically complex and can present at various times during treatment, including immediately after treatment initiation (primary resistance), weeks or months after evidence of the initial clinical benefit (secondary resistance), or after treatment has been halted for a variety of reasons^{9,25}.

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| Studies | Population | Population Histology | EGFR-TKIs | ORR, % | ORR, % Median OS, Median PFS, | Median PFS, | PI | Progression-free survival | rival |
|-------------------------------|------------|----------------------|-------------------|--------|-------------------------------|-------------|-----------------|-----------------------------------|-----------------|
| | | | | | months | months | 3 months, n (%) | 3-6 months, n (%) 6 months, n (%) | 6 months, n (%) |
| Goto, 2013 ³⁵ | Japanese | ADC (n=102) | Erlotinib (n=102) | 78.0 | NA | 11.8 | 13 (12.7) | 17 (16.7) | 72 (70.6) |
| Noronha, 2020 ¹³ | Indian | ADC (n=170) | Gefitinib (n=176) | 62.5 | 17.0 | 8.0 | 20 (11.4) | 23 (13.1) | 33 (75.5) |
| | | ADS (n=4) | | | | | | | |
| Wiii 2018 ¹⁴ | Chinese | ADC (n=217) | Afatinih (n-217) | 66 A | 23.1 | 11.0 | 32 (14 7) | 30 (18 0) | 146 (67 3) |
| Dovis 2016 ³⁶ | Vor.000 | | | 0.00 | 0.20 | | | 00 (10 0) | |
| raik, zuio | NOLEAL | | | 0.07 | 21.3 | 0.11 | (7.11) 01 | (0.01) 00 | (0.07) 211 |
| | | ADS (n=2) | Gefitinib (n=159) | 56.0 | 25.0 | 10.9 | 27 (17.0) | 26 (16.3) | 106 (66.7) |
| Pluzanski, 2020 ¹⁵ | Polish | ADC (n=619) | Erlotinib (n=252) | 53.8 | 20.4 | 12.1 | 36 (14.3) | 55 (21.8) | 161 (63.9) |
| | | | Gefitinib (n=255) | 56.5 | 17.5 | 10.3 | 38 (14.9) | 57 (22.4) | 160 (62.7) |
| | | | Afatinib (n=112) | 50.0 | NA | 16.4 | 14 (12.5) | 38 (33.9) | 60 (53.6) |
| Kuan, 2016 ³⁷ | Taiwanese | ADC (n=448) | Erlotinib (n=63) | NA | NA | 11.2 | 10 (15.9) | 9 (14.3) | 44 (69.8) |
| | | | Gefitinib (n=304) | | | 12.1 | 34 (11.2) | 51 (16.8) | 219 (72.0) |
| | | | Afatinib (n=81) | | | 10.3 | 4 (4.9) | 12 (14.8) | 65 (80.3) |

EGFR-TKIs=epidermal growth factor receptor tyrosine kinase inhibitors, NSCLC=non-small cell lung cancer, ADC=adenocarcinoma, ADS=adenosquamous carcinoma, SCC=squamous cell carcinoma, PFS=progression free survival, ORR=objective response rate, OS=overall survival, NA=not available

Primary resistance

Primary resistance is defined as a failure to respond to treatment for the first time after receiving EGFR-TKIs, without any evidence of an objective response. Primary resistance in NSCLC patients can be divided into 4 classes: 1) harboring drug-resistant EGFR mutations; 2) genomic alterations along with drug-sensitizing EGFR mutations; 3) EGFR wild-type; and 4) unknown mechanisms²⁶. The most prevalent mutations that result in primary resistance to EGFR-TKIs in drug-resistant EGFR mutation are the T790M mutation and insertions in exon 20²⁷. In addition, many studies have shown that genomic alterations, along with drug-sensitizing EGFR mutations, may be linked to primary resistance. These details are summarized in Table 3. Clinical samples, including tissue and blood, were obtained prior to treatment in patients treated with either of the EGFR first- or second-generation TKIs (gefitinib, erlotinib, afatinib, and icotinib).

The clinical definitions of primary resistance provided are diverse and non-uniform. Two studies define primary resistance to EGFR-TKIs as a combination of no evidence of objective response rate (ORR) and progression-free survival (PFS) for less than 3¹¹ and 4 months²⁸, respectively. In contrast, the other 2 studies only considered PFS of less than 6 months as the criterion for primary resistance^{29,30}. The genomic alteration of all clinical samples was determined using target sequencing, which involved analyzing a set of 22-458 genes. The TP53 mutation was the most common genetic alteration reported in all of the studies. It was higher in cases conferring primary resistance than responders: 100.0% vs. 45.7%²⁸, 80.0% vs. 52.9%, and 87.5% vs. 16.7%³⁰. Four genes were found in 3 studies exhibiting different mutation frequencies. CTNNB was found in 11.1% of patients with primary resistance¹¹, 5.0% in responders vs. 5.2% in non-responders²⁸, and 13.3% in short PFS vs. 17.6% in long PFS. SMAD4 was found in 11.1% of patients with primary resistance¹¹, 10% of nonresponders vs 23.3% of non-responders²⁸, and 33.3% in short PFS²⁹. PTEN was found in 5% of non-responders vs 1.7% of non-responders²⁸, 5.9% in long PFS²⁹, and 6.3% in short-term responders³⁰. ERBB2-4 was found in 5–15% of non-responders vs 6.0–8.6% in responders²⁸, 6.7% in short PFS vs. 5.9% in long PFS²⁹, and 12.5% in short-term responders³⁰. In addition, a study conducted by Jin et al. (2018) has shown that RB1, which is only found in the short PFS, has a high mutation frequency, accounting for 26.7% of cases. In contrast, another study by Xu *et.al.* (2018) observed that the frequency of RB1 mutation was comparable in both short-term responders (31.3%) and long-term responders (33.3%) (Table 3).

Acquired resistance

EGFR-TKIs have been shown to have good response rates and prolong PFS in NSCLC patients with sensitive activating EGFR mutations. Despite the initial benefits, almost all patients will experience significant tumor progression or acquire resistance to EGFR-TKI. The clinical definition of acquired resistance is non-uniform. Jackman et al. (2010) have proposed a criteria for defining more precisely acquired resistance to EGFR-TKIs. This criteria consists of: 1) previously received treatment with a singleagent EGFR-TKI (first- or second-generation); 2) either or both of the following: a tumor that harbors an EGFR mutation known to be associated with drug sensitivity or objective clinical benefit from treatment with an EGFR-TKI (either partial/complete response or stable disease longer than 6 months. evaluated by RECIST or WHO); 3) systemic progression of disease while on continuous treatment with EGFR-TKI within the last 30 days; and 4) no intervening systemic therapy between cessations of EGFR-TKI and the initiation of a new therapy¹⁰.

It is well known that a secondary point mutation that substitutes methionine for threonine at amino acid position 790 (T790M) is mainly the molecular mechanism Table 3 Genomic alterations in EGFR-mutant NSCLC with primary resistance to EGFR-TKI

| Study | Population | Sample/Drug | Sample collection/Number | Genomic alteration |
|-------------------------|------------|--|---|--|
| Lee, 2013 ¹¹ | Korean | - FFPE - Gef (n=179) - Erl (n=18) | Definition of primary resistance: - PFS <3 months - no evidence of ORR - 11 cases (11/197, 5.6%) | TP53 (3/9, 33.3%)/NOTCH1 (1/9, 11.1%)/CTNNB1 (1/9, 11.1%)/SMAD4 (1/9, 11.1%) Note: Sequence only in cases of primary resistance |
| Lim, 2016 ²⁸ | Korean | - FFPE - Gef (n=136) | Definition of primary resistance: - PFS <4 months - no evidence of ORR - 20 cases (20/136, 14.7%) | Non-responders: TP53 (20/20, 100%)/DDR2, MET, PIK3CA, STK11 (4/20, 20%)/ERBB4, FGFR1-3, KRAS (3/20, 15%)/AKT1, ALK, MAP2K1, SMAD4 (2/20, 10%)/BRAF, CTNNB1, ERBB2, FBXW7, PTEN (1/20, 5%) Responders: TP53 (53/16, 45.7%)/ SMAD4 (27/116, 23.3%)/ DDR2 (23/116, 19.8%)/ STK11 (18/116, 15.5%)/ MET (14/116, 12.1%)/ FGFR1-3 (11/116, 9.5%)/ ERBB4 (10/116, 8.6%)/BRAF (9/116, 7.8%)/ FBBB2 (8/116, 6.9%)/AKT1, PIK3CA (7/116, 6.0%)/ MAP2K1, NFAS, CTNNB1 (6/116, 5.2%)/ FBXW7 (5/116, 4.3%)/ NOTCH1, ALK (3/116, 2.6%)/ PTEN (2/116, 1.7%) |
| Jin, 2018 ²⁸ | Chinese | - FFPE/plasma - Gef (n=17) - Ico (n=51) - Afa (n=1) | Definition of primary resistance: - PFS <6 months (Short PFS; n=15) - PFS >12 months (Long PFS; n=17) | Short PES: TF53 (12/15, 80.0%)/SMAD4 (5/15, 33.3%)/RB1 (4/15, 26.7%)/ARID14, PIK3CA (3/15, 20.0%)/CDC73, EP300, MCL1, AKT1, CTNNB1, STK11 (2/15, 13.3%)/TSHR, CDKN2B, CREBBP, JAK2, MLH1, MAP2K4, FGFR3, HGF, NRAS, PRKCI, CCNE1, PRF1, NKX2-1, ARAF, JUN, NBN, RICTOR, ARID2, FLT1, POT1, NOTCH2, DNMT3A, APC, EPHA3, ERBB3, FAT1, BLM, PDE114, ROS1, PDK1, PT7, ATR, PMS2, NFKBIA (1/16, 6.7%) Long PES: TP53 (9/17, 52.9%)/CTNNB1, PDK1 (3/17, 17.6%)/AKT1, FLT4, THADA (2/17, 11.8%)/AKT2, CDK6, TSHR, SMAD2, CTCF, PTEN, EP300, PIK3CA, MLH1, ERCC4, CHD4, CSF1R, GRIN2A, NF1, PPP2R1A, GATA1, PGR, NKX2-1, MYC, ARAF, JUN, NSD1, KDM5A, KMT2A, RAD51D, POT1, NOTCH2NL, CYP3A4, APC, EPHA2, ERBB3, FGFR4, SOX2, BUB1B, PDE11A,RAD50, PDCD1, ATRX, POLE, MDM2, RECQL4, SMARCA4, TNFSF11, TOP2A, YAP1 (1/17, 5.9%) Note: Only found in Short PFS and confer primary resistance. SMAD4, RB1, ARID1A, CDC73, STK1, MCL1, ARID2, DNMT3A, ATR, ARAF, HGF, PRKCI, CNE1, ARAF |
| Xu, 2018 ³⁰ | Chinese | - FFPE - Gef (n=11) - Ico (n=16) - Erl (n=1) | Definition of primary resistance: - PFS <6 months (Short-term responders; n=16) - PFS >24 months (Long-term responders; n=12) | Short-term responders: TP53 (14/16, 87.5%)/EGFR amp. (6/16, 37.5%)/RB1 (5/16, 31.3%)/ ERBB2 (2/16, 12.5%)/PTEN, MET, AKT2, HRAS (1/16, 6.3%)/Mo mutation in AKT2 Long-term responders: RB1 (4/12, 33.3%)/TP53, AKT1 (2/12, 16.7%)/EGFR amp. (1/12, 8.3%)/ No mutation in PTEN, ERBB2, MET, AKT2, HRAS |

Table 4 Genomic alterations in EGFR-mutant NSCLC with acquired resistance to EGFR-TKI

| Study | Population | Sample/Drug | Sample collection/Number | Genomic alteration |
|----------------------------|-------------|--|--|--|
| Jin, 2016 ³⁴ | Chinese | - FFPE/pleural effusions - Gef (n=26) - Erl (n=15) - Ico (n=42) | - Post-treatment - T790M+; 30 cases - T790M-; 53 cases | T790M±: multiple EGFR (30/30, 100.0%)/TP53 (21/30, 70.0%)/TET1 (4/30, 13.3%)/APC, RB1 (2/30, 6.7%)/ERBB2, SOX2 (1/30, 3.3%)/No mutation in KRAS, MET, ALK, FGFR1 T790M-: multiple EGFR (26/53, 49.1%)/TP53 (27/53, 50.9%)/ TET1 (6/53, 11.3%)/ ERBB2 (5/53, 9.4%)/ KRAS, MET, SOX2 (4/53, 7.5%)/APC, RB1, ALK, FGFR1 (3/53, 5.7%) |
| Lee, 2017 ³² | Korean | FFPE/pleural effusions Gef (n=17) Erl (n=2) | Pre-treatment and post-treatment T790M+; 30 cases T790M-; 12 cases | T790M+: SYNE1 (2/12, 16.7%)/TGA10, BCL2L2, TGM7, MSH2, RET, MAGEA1, PIK3CG, LPHN3, CREBBP, CMPK1, NTRK3, GRM8, NOTCH4, PLCG1, AMER1, MAGEA1, MN1, DST ² , TAF1 ² , EANCA ² , KAT6B (1/12, 8.3%) T790M-: TP53 ² (3/5, 60.0%)/ CSMD3 ² , AKAP9 (2/5, 40.0%)/SEPT9, EML4, MUTYH, AKT3, ATR, LIFR, ATRX, SOX2, CD79B, AXL, COL1A1, MAML2, CYP2D6, KMT2D, XPC, JAK2, MAF, AR ² , CASC5, FGFR4, ERCC2, RAF1, PBX1, CTNNB1, NIN, NUP214, MYH9, NUP98, PDE4DIP, KDM5C, RNF213, RNASEL, TOP1, TGFBR2, TRIM24, IGF1R (1/5, 20.0%) Note: [*] is significantly mutated genes in post-treatment compared to pre-treatment |
| Otsubo, 2019 ³¹ | Japanese | - Plasma - Gef (n=5) - Erl (n=7) - Afa (n=12) | - Post-treatment - T790M+; 17 cases - T790M-; 7 cases | T790M+: TP53 (4/7, 57.1%)/MET (3/7, 42.9%)/ERBB2 (1/7, 14.3%) T790M-: MET (6/17, 35.3%)/TP53, PIK3CA, APC (5/17, 29.4%), multiple EGFR (4/17, 23.5%) |
| Chua, 2021 ³⁸ | Singaporean | - Fresh-frozen tissue - 1 st and 2 nd EGFR TKI (n=59) | - Post-treatment - T790M+; 38 cases - T790M-; 21 cases | T790M+: TP53 (19/38, 50%)/FOXA1 (9/38, 24%)/amplification EGFR, NKX2- 1, YEATS4, CDH10 (8/38, 21.0%)/MDM2, PIK3CA (7/38, 18%)/Arm Chr3q, FGFR4 (5/38, 13%)/RB1, CTNNB1 (3/38, 8%)/FEBB2, PTEN, HGF (2/38, 5%)/MET, AXIN1 (1/38, 3%)/ No mutation in PIK3CB T790M-: TP53 (18/21, 86%)/ Arm Chr3q (12/21, 57%)/MET, HGF (4/21, 19%)/ amplification EGFR, PIK3CA (3/21, 14%)/ ERBB2, RB1, PIK3CB (2/21, 10%)/ MDM2, PTEN, FOXA1, NKX2-1, YEATS4 (1/21, 5%)/No mutation in FGFR4, APC, CTNNB1, AXIN1, CDH10 |

-empeaaea, bar IIXed Ó -טמווו E B H mutation, EGFR-mutant NSCLC=non-small cell lung cancer with EGF T790M+=T790M mutation, T790M-=non-T790M mutation that produces acquired resistance after drug treatment. In addition, concurrent genomic alterations in genes other than EGFR may be responsible for acquired resistance (Table 4). Clinical samples, including tissues, pleural effusions and blood, were obtained at the time of acquired resistance to EGFR-TKI (gefitinib, erlotinib, afatinib, and icotinib). All studies have shown that patients having received EGFR-TKI develop the T790M mutation (T790M+) at: 70.8% by Otsubo et al. (2019)³¹, 71.4% by Lee et al. (2017)³², 64.4% by Chua et al. (2021)³³, and 36.1% by Jin et al. (2016)³⁴. In addition, genetic alterations coexisting with T790M, as well as concurrent genomic alterations in genes other than EGFR, may be responsible for acquired resistance. TP53 was the most frequently mutated gene across all the studies, 2 of which showed both T790M+ and non-T790M- mutation (T790M-)^{33,34}, whereas in the other 2 studies, it was only shown to be present in T790M+³¹ or T790M-³². In the T790M+, the mutation of multiple EGFR positions (100.0%)³⁴ and MET (42.9%)³¹ was found in high frequencies. Similarly, high mutation frequencies have been identified at multiple EGFR positions (49.1%)³⁴, CSMD3 (40.0%)³², AKAP9 (40.0%)³², MET (35.3%)³¹ and Arm Chr3q $(57\%)^{33}$.

Conclusion

First or second-generation EGFR-TKIs are the approved first-line therapy for NSCLC with EGFR-activating mutations. This report provides a comprehensive overview of many studies evaluating the effectiveness of EGFR-TKI treatments in various populations of advanced NSCLC patients. The studies evaluated various outcomes, including ORR, OS and PFS. We additionally reviewed concurrent gene alterations related to both primary and acquired resistance. Our findings could aid in the identification of patients that may derive more benefits from EGFR-TKI treatment, even though possessing sensitive EGFR mutations.

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Conflict of interest

The authors declare no conflict of interest.

References

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin 2021;71:209–49.
- Rojanamatin J, Ukranun W, Supaattagorn P, Chaiwiriyabunya I, Wongsena M, Chaiwerawattana A, et al. Cancer in Thailand volume X 2016–2018. Bangkok: National Cancer Institute Ministry of Public Health; 2021.
- Sher T, Dy GK, Adjei AA. Small cell lung cancer. Mayo Clinic proceedings 2008;83:355–67.
- Saito M, Suzuki H, Kono K, Takenoshita S, Kohno T. Treatment of lung adenocarcinoma by molecular-targeted therapy and immunotherapy. Surg Today 2018;48:1–8.
- Shi K, Wang G, Pei J, Zhang J, Wang J, Ouyang L, et al. Emerging strategies to overcome resistance to third-generation EGFR inhibitors. J Hematol Oncol 2022;15:94.
- Zhou C, Wu YL, Chen G, Feng J, Liu XQ, Wang C, et al. Erlotinib versus chemotherapy as first-line treatment for patients with advanced EGFR mutation-positive non-small-cell lung cancer (OPTIMAL, CTONG-0802): a multicentre, open-label, randomised, phase 3 study. Lancet Oncol 2011;12:735-42.
- Sharma J, Shum E, Chau V, Paucar D, Cheng H, Halmos B. The evolving role of biomarkers in personalized lung cancer therapy. Respiration 2017;93:1–14.
- Karachaliou N, Fernandez-Bruno M, Bracht JWP, Rosell R. EGFR first- and second-generation TKIs-there is still place for them in EGFR-mutant NSCLC patients. Transl Cancer Res 2019;8(Suppl1):S23-47.
- Ayoola A, Barochia A, Belani K, Belani CP. Primary and acquired resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-small cell lung cancer: an update. Cancer Invest 2012;30:433–46.

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- Jackman D, Pao W, Riely GJ, Engelman JA, Kris MG, Janne PA, et al. Clinical definition of acquired resistance to epidermal growth factor receptor tyrosine kinase inhibitors in non-smallcell lung cancer. J Clin Oncol 2010;28:357–60.
- Lee JK, Shin JY, Kim S, Lee S, Park C, Kim JY, et al. Primary resistance to epidermal growth factor receptor (EGFR) tyrosine kinase inhibitors (TKIs) in patients with non-small-cell lung cancer harboring TKI-sensitive EGFR mutations: an exploratory study. Ann Oncol 2013;24:2080–7.
- Guo Y, Song J, Wang Y, Huang L, Sun L, Zhao J, et al. Concurrent genetic alterations and other biomarkers predict treatment efficacy of EGFR-TKIs in EGFR-mutant non-small cell lung cancer: a review. Front Oncol 2020;10:610923.
- Noronha V, Patil VM, Joshi A, Menon N, Chougule A, Mahajan A, et al. Gefitinib versus gefitinib plus pemetrexed and carboplatin chemotherapy in EGFR-mutated lung cancer. J Clin Oncol 2020;38:124–36.
- Wu YL, Xu CR, Hu CP, Feng J, Lu S, Huang Y, et al. Afatinib versus gemcitabine/cisplatin for first-line treatment of Chinese patients with advanced non-small-cell lung cancer harboring EGFR mutations: subgroup analysis of the LUX-Lung 6 trial. Onco Targets Ther 2018;11:8575–87.
- 15. Pluzanski A, Krzakowski M, Kowalski D, Dziadziuszko R. Real-world clinical outcomes of first-generation and secondgeneration epidermal growth factor receptor tyrosine kinase inhibitors in a large cohort of European non-small-cell lung cancer patients. ESMO Open 2020;5:e001011.
- Hodoglugil U, Carrillo MW, Hebert JM, Karachaliou N, Rosell RC, Altman RB, et al. PharmGKB summary: very important pharmacogene information for the epidermal growth factor receptor. Pharmacogenetics and genomics. 2013;23:636–42.
- Richards CS, Bale S, Bellissimo DB, Das S, Grody WW, Hegde MR, et al. ACMG recommendations for standards for interpretation and reporting of sequence variations: Revisions 2007. Genet Med 2008;10:294–300.
- Kaler AK, Patel K, Patil H, Tiwarekar Y, Kulkarni B, Hastak M, et al. Mutational Analysis of EGFR Mutations in Non-Small Cell Lung Carcinoma-An Indian Perspective of 212 Patients. Int J Environ Res Public Health 2022;20.
- Tanaka T, Matsuoka M, Sutani A, Gemma A, Maemondo M, Inoue A, et al. Frequency of and variables associated with the EGFR mutation and its subtypes. Int J Cancer 2010;126:651–5.

- Midha A, Dearden S, McCormack R. EGFR mutation incidence in non-small-cell lung cancer of adenocarcinoma histology: a systematic review and global map by ethnicity (mutMapII). Am J Cancer Res 2015;5:2892–911.
- Levin PA, Mayer M, Hoskin S, Sailors J, Oliver DH, Gerber DE. Histologic transformation from adenocarcinoma to squamous cell carcinoma as a mechanism of resistance to EGFR Inhibition. J Thorac Oncol 2015;10:e86–8.
- Zubair T, Bandyopadhyay D. Small molecule EGFR inhibitors as anti-cancer agents: discovery, mechanisms of action, and opportunities. Int J Mol Sci 2023;24.
- Abourehab MAS, Alqahtani AM, Youssif BGM, Gouda AM. Globally approved EGFR inhibitors: insights into their syntheses, target kinases, biological activities, receptor interactions, and metabolism. Molecules (Basel, Switzerland) 2021;26.
- Girard N. Optimizing outcomes in EGFR mutation-positive NSCLC: which tyrosine kinase inhibitor and when? Future oncology (London, England) 2018;14:1117–32.
- Simasi J, Schubert A, Oelkrug C, Gillissen A, Nieber K. Primary and secondary resistance to tyrosine kinase inhibitors in lung cancer. Anticancer Res 2014;34:2841–50.
- Sharma SV, Bell DW, Settleman J, Haber DA. Epidermal growth factor receptor mutations in lung cancer. Nat Rev Cancer 2007;7:169–81.
- VanderLaan PA, Rangachari D, Mockus SM, Spotlow V, Reddi HV, Malcolm J, et al. Mutations in TP53, PIK3CA, PTEN and other genes in EGFR mutated lung cancers: correlation with clinical outcomes. Lung Cancer 2017;106:17–21.
- Lim SM, Kim HR, Cho EK, Min YJ, Ahn JS, Ahn MJ, et al. Targeted sequencing identifies genetic alterations that confer primary resistance to EGFR tyrosine kinase inhibitor (Korean Lung Cancer Consortium). Oncotarget 2016;7:36311–20.
- Jin Y, Shi X, Zhao J, He Q, Chen M, Yan J, et al. Mechanisms of primary resistance to EGFR targeted therapy in advanced lung adenocarcinomas. Lung Cancer 2018;124:110–6.
- 30. Xu Y, Tong X, Yan J, Wu X, Shao YW, Fan Y. Short-Term responders of non-small cell lung cancer patients to EGFR tyrosine kinase inhibitors display high prevalence of TP53 mutations and primary resistance mechanisms. Transl Oncol 2018;11:1364–9.
- Otsubo K, Sakai K, Takeshita M, Harada D, Azuma K, Ota K, et al. Genetic profiling of non-small cell lung cancer at development of resistance to first- or second-generation EGFR-TKIs by

CAPP-Seq analysis of circulating tumor DNA. The Oncologist 2019;24:1022-6.

- 32. Lee CK, Kim S, Lee JS, Lee JE, Kim SM, Yang IS, et al. Nextgeneration sequencing reveals novel resistance mechanisms and molecular heterogeneity in EGFR-mutant non-small cell lung cancer with acquired resistance to EGFR-TKIs. Lung Cancer 2017;113:106-14.
- Chua KP, Teng YHF, Tan AC, Takano A, Alvarez JJS, Nahar R, et al. Integrative profiling of T790M-negative EGFR-mutated NSCLC reveals pervasive lineage transition and therapeutic opportunities. Clin Cancer Res 2021;27:5939–50.
- 34. Jin Y, Shao Y, Shi X, Lou G, Zhang Y, Wu X, et al. Mutational profiling of non-small-cell lung cancer patients resistant to first-generation EGFR tyrosine kinase inhibitors using next generation sequencing. Oncotarget 2016;7:61755–63.
- 35. Goto K, Nishio M, Yamamoto N, Chikamori K, Hida T, Maemondo M, et al. A prospective, phase II, open-label study (JO22903) of first-line erlotinib in Japanese patients with epidermal growth factor receptor (EGFR) mutation-positive advanced non-small-cell lung cancer (NSCLC). Lung Cancer 2013;82:109-14.
- Park K, Tan EH, O'Byrne K, Zhang L, Boyer M, Mok T, et al. Afatinib versus gefitinib as first-line treatment of patients with EGFR mutation-positive non-small-cell lung cancer (LUX-Lung 7): a phase 2B, open-label, randomised controlled trial. Lancet Oncol 2016;17:577–89.
- Kuan FC, Li SH, Wang CL, Lin MH, Tsai YH, Yang CT. Analysis of progression-free survival of first-line tyrosine kinase inhibitors in patients with non-small cell lung cancer harboring leu858Arg or exon 19 deletions. Oncotarget 2017;8:1343–53.